

Evidence of NF-κB pathway activation in patients with advanced, high molecular risk myelofibrosis

Jennifer O'Sullivan, MD^{1,3*}, Aaron T. Gerds, MD, MS,² Claire N. Harrison, DM,³ Stephen T. Oh, MD,⁴ Angela Hamblin, MD, PhD,⁵ Sarah A Buckley, MD,⁶ Alan F. List, MD⁷ and Adam J. Mead, MBBChir^{1,5}

¹MRC Weatherall Institute of Molecular Medicine, University of Oxford, Oxford, United Kingdom; ²Cleveland Clinic Taussig Cancer Institute, Cleveland, OH; ³Guy's and St Thomas' NHS Foundation Trust, London, United Kingdom; ⁴Washington University School of Medicine, Saint Louis, MO; ⁵NIHR Biomedical Research Centre, University of Oxford, Oxford, United Kingdom; ⁶CTI BioPharma, Seattle, WA; ⁷Precision BioSciences, Inc., Durham, NC



INTRODUCTION

- Patients with myelofibrosis who discontinue treatment with the JAK1/2 inhibitor ruxolitinib have a poor prognosis that is often associated with advanced phases of disease and severe cytopenias.
- PAC203, a Phase 2 dose-finding study in patients with symptomatic myelofibrosis who were intolerant of or resistant to ruxolitinib¹, represents an opportune cohort for analyzing the association between high molecular risk (HMR) mutations and disease phenotype.
- While these patients are more likely to have high molecular risk genomic markers, biological drivers of disease in this advanced population are not well characterized. Pro-inflammatory cytokines have been shown to be elevated in MF² but no associations with mutation profiles have been identified.
- Patients had advanced disease at study entry, with profound cytopenias and high mutational burden³.
- Here, we analyzed the interaction between high-risk mutations and cytokine profiles of patients treated in PAC203.

OBJECTIVE

- To study the interaction between high-risk mutations and cytokine profiles of patients treated in the PAC203 study.

METHODS

- Trial entry cytokine and mutation data were available in 108 (of total 164 recruited; 161 treated) patients.
- Using the Myriad RBM platform, a microsphere-based immuno-multiplexing technology, 47 cytokines were assessed.
- Mutation profiles were determined using an ISO accredited Illumina TruSeq Custom Amplicon Panel, including 32 gene mutation hotspots and exons (~36,000 bp, 287 amplicons). CALR mutation screening was carried out independently. Accepted coverage was achievement of a depth of ≥100 reads per base in ≥95% of targeted bases.
- The initial analysis assessed possible relationships between individual plasma cytokine levels and somatic gene mutation and clinical demographic data.
- An unsupervised approach was then used applying hierarchical agglomerative clustering to identify related sets of cytokines.
- Cluster scores (based on the median overall cytokine concentration within each cluster for each patient) were correlated with clinical/genomic data.

Clinical and molecular patient characteristics

- The PAC203 cohort has a high prevalence of severe thrombocytopenia and anemia (**Table 1**).
- The mutation profile of patients included for this analysis had a mutation profile enriched for high molecular risk (HMR) mutations (*IDH1/2*, *SRSF2*, *ASXL1*, *EZH2*, *U2AF1Q1574*) as previously described³ (**Table 1**).
- In addition, splicing factor mutations (SF; *SF3B1*, *U2AF1*, *ZRSR2*, *SRSF2*) were detected in 35% of patients (**Table 1**).
- RAS-pathway activating mutations (*KRAS/NRAS/CBL*) mutations were present at a higher frequency than reported to date in any other MF cohorts^{5,6} (**Table 1**).

Table 1. Patient Demographics

Demographics	Patients (n=108)
Follow-up time (median, range in days)	210 (27 – 526)
Age (median, range in years)	68 (37 - 87)
Primary MF (n,%)	61 (56.5)
Platelet count x 10 ⁹ /L (median, range)	64 (13 – 910)
Platelet count <50 x 10 ⁹ /L (n,%)	41 (38.3)
Hemoglobin level <10g/dL (n,%)	69 (64.5)
JAK-STAT signalling driver mutations (n,%)	
JAK2V617F	84 (77.8)
CALR	14 (12.9)
MPL	8 (7.4)
Triple negative	2 (1.9)
Addition somatic driver mutations (n,%)	
≥3 additional somatic driver mutations	21 (19.4)
High molecular risk	44 (40.7)
Splicing factor	35 (32.4)
RAS-pathway activating	23 (21.3)
ASXL1	29 (26.9)
TET2	26 (24.1)
TP53	7 (6.5)

Cytokine analysis

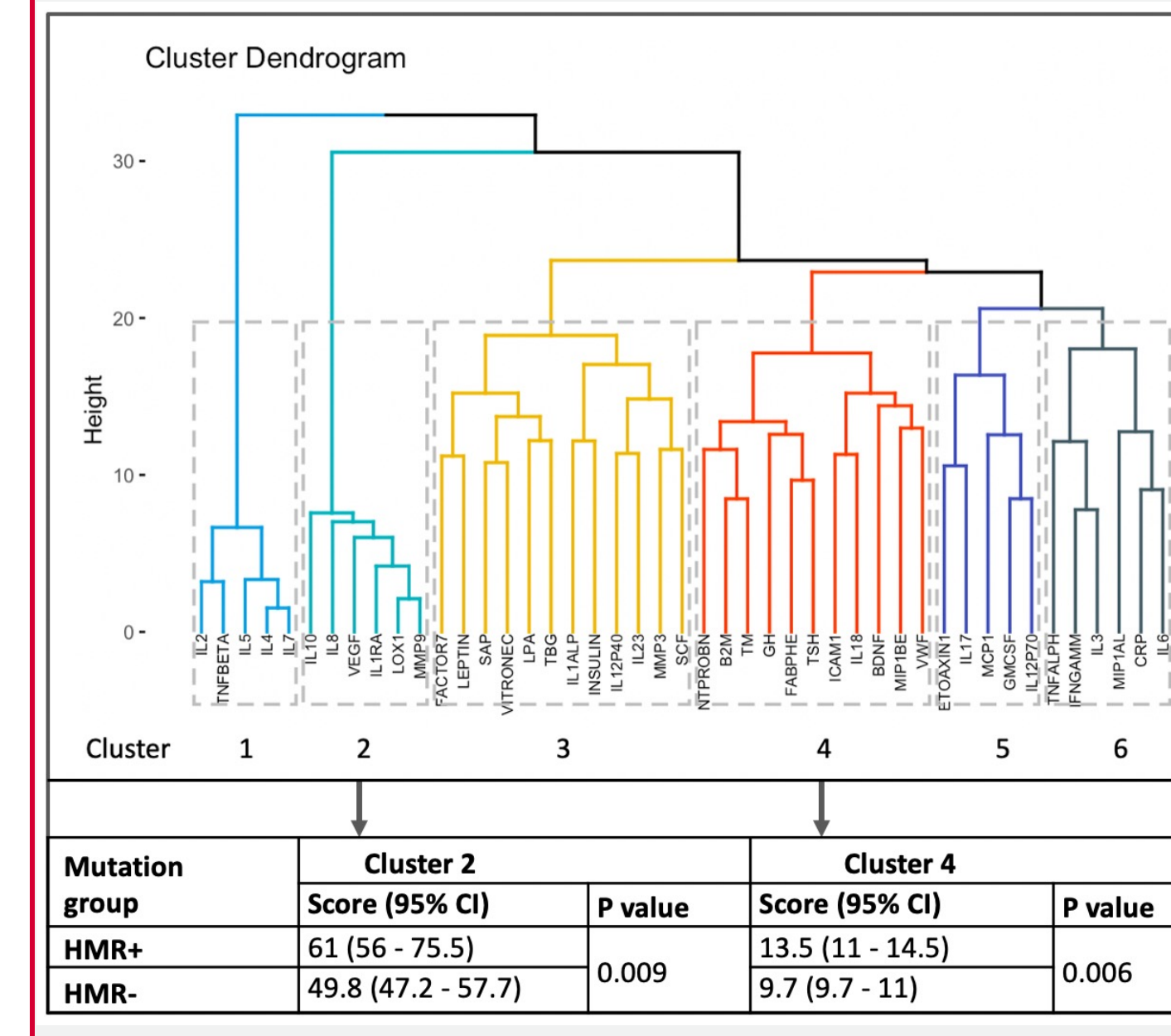
- Unsupervised agglomerative clustering to identify sets of related cytokines identified 6 clusters (**Figure 1**).
- Clusters 2, 4, 5 and 6 are enriched for pro-inflammatory markers, with cluster 2 representing a transcriptional cluster regulated by the NF-κB pathway (interleukin-8 [IL-8], IL-10, IL-1 receptor alpha, vascular endothelial growth factor [VEGF]).

RESULTS

Cytokine-mutation correlations

- Elevations in clusters 2 and 4 were associated with presence of HMR mutations (HMR+).
 - Cluster 2 in particular represents a set of cytokines regulated by NF-κB
- Within cluster 2, IL-8 levels were most strongly associated with HMR status (HMR+, 40.5 pg/ml vs. HMR-, 24.5 pg/ml, $P < 0.0001$).

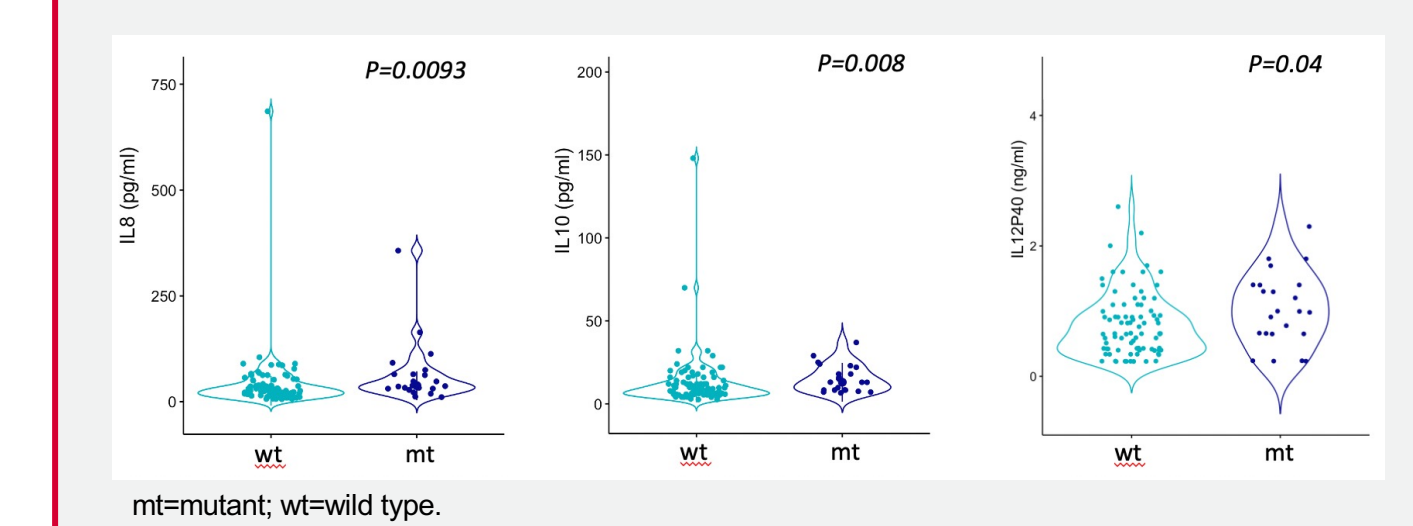
Figure 1. Cluster dendrogram of cytokine levels with cluster scores 2 and 4 highlighted in the table for high molecular risk positive (HMR+) and negative (HMR-) patients.



- Although cluster 6 did not correlate with HMR status, it did correlate with presence of the high-risk *U2AF1* mutation (mutated 19.3 vs. unmutated 6.3, $P = 0.0019$), though not with other splicing factor mutations. Furthermore, levels of the cluster 6 cytokine tumour necrosis factor-alpha (TNF-α) were associated with HMR (HMR+ 61 pg/mL vs. HMR- 49 pg/mL, $P = 0.0088$).
- No significant associations between cluster scores and driver mutations or clinical characteristics were identified.

- RAS-pathway mutations were associated with higher levels of the NF-κB-associated cytokines IL-8, IL-10, and IL-12P40 (**Figure 2**), though not with overall cluster 2 scores.

Figure 2. Levels of individual cytokines elevated in patients with RAS-activating mutations



- For RAS-mt vs. RAS-wt, median cytokine levels comparisons were for IL-8 (38 pg/ml vs. 26 pg/ml, $P = 0.0093$), IL-10 (13 pg/ml vs. 9.1 pg/ml, $P = 0.008$) and IL12P40 (1 ng/ml vs 0.6 ng/ml, $P = 0.04$) (**Figure 2**).
- There was no association between cytokine cluster scores and recent exposure to ruxolitinib.

CONCLUSIONS

- We report associations between high-risk mutation profiles and cytokine signatures in patients with myelofibrosis.
- In this HMR+ and RAS mutant-enriched cohort of MF patients who were intolerant of or resistant to ruxolitinib, we identify a relationship between HMR mutations and an NF-κB directed pro-inflammatory cytokine signature.
- These results implicate the activation of a distinct biological signalling pathway operative in this molecularly-defined cohort

REFERENCES: 1. Gerds AT, Savona MR, Scott BL, et al. *Blood Advances*. 2020;4(22):5825-5835. 2. Tefferi A, Vaidya R, Caramazza D, et al. *J Clin Oncol*. 2011;29:1356-1363. 3. O'Sullivan J et al. *Blood* (2019) 134 (Sup_1):4214. 4. Tefferi et al. *J Clin Oncol*. 2018;36(17):1769-1770. 5. Coltro G, Rotunno G, Mannelli L, et al. *Blood Advances*. 2020;4(15):3677-3687. 6. Santos FPS, Getta B, Masarova L, et al. *Leukemia*. 2020;34(3):799-810.

ACKNOWLEDGEMENTS: Supported in part by a Medical Research Council Senior Clinical Fellowship (AJM, MR1006340/1), a Cancer Research UK (CRUK) Senior Cancer Research Fellowship (AJM) and by the NIHR Oxford Biomedical Research Centre based at Oxford University Hospitals NHS Trust and University of Oxford (AJM).

QR Code
FPO

*Presenting Author: Jennifer O'Sullivan; jennifer.osullivan@rdm.ox.ac.uk